

11-29-00

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PATENT  
674544-2001

**Patent Application Transmittal**  
**(only for new nonprovisional applications**  
**under 37 C.F.R. 1.53(b))**  
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JC520 U.S. PTO  
09/724985  
11/28/00

Date: November 28, 2000

TO: ASSISTANT COMMISSIONER FOR PATENTS  
Box PATENT APPLICATION  
Washington, D.C. 20231

Sir:

With reference to the filing in the United States Patent and Trademark Office of an application for patent in the name of:

**Philip Michael SAVAGE**

Entitled: **METHOD FOR PRODUCING OR ENHANCING A T-CELL RESPONSE  
AGAINST A TARGET CELL USING A COMPLEX COMPRISING AN  
HLA CLASS 1 MOLECULE AND AN ATTACHING MEANS**

- This is an application of a small entity under 37 CFR 1.9(f) and the amounts shown in parentheses below have been employed in calculating the fee.
- Undersigned attorney hereby states that this is an application of a Small Entity.

The following are enclosed:

- X   Specification (28 pages) (ONE PAGE ABSTRACT)
- X   9 Sheet(s) of Drawings (Figs. 1-9)
- X     28   Claims (including 1 independent claim)
- This application contains multiple dependent claim(s)
- X   A Petition To Make Special.
- X   Our checks totaling \$1,024.00, calculated on the basis of the claims as amended by any enclosed preliminary amendment as follows:

JMM3529

Basic Fee  
\$710.00 (355.00).....710.00

Total Number of Claims in excess  
of 20 at \$18.00 (9.00) each .....144.00

Number of Independent Claims in  
excess of 3 at \$80.00 (40.00) each.....000.00

Multiple Dependent Claim Fee  
at \$270.00 (135.00).....000.00

Total Filing Fee.....854.00

Assignment Recording Fee \$40.00 ..... 40.00

Petition to Make Special Fee \$130.00 .....130.00

  X   Oath or Declaration and Power of Attorney

       New        signed   X   unsigned  
       Copy from prior application (37 C.F.R. 1.63(d))

       Certified copy of the following application to substantiate the claim for priority  
made in the Declaration:

Please charge any additional fees required for the filing of this application or credit any  
overpayment to Deposit Account No. 50-0320. Please accord this application an November 22,  
2000 filing date.

Respectfully submitted,

FROMMER LAWRENCE & HAUG LLP

By: Thomas J. Kowalski  
Thomas J. Kowalski  
Reg. No. 32,147  
(212) 588-0800

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November 28, 2000

Assistant Commissioner for Patents  
Washington, D.C. 20231

Re: Continuation-in-Part of PCT Application – 35 U.S.C. 111(a)  
Applicant: Philip Michael SAVAGE  
Our Ref: 674544-2001

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Dear Sir:

Enclosed are papers constituting the above patent application which is being filed under 37 C.F.R. 1.53 as a **Continuation-in-Part of a PCT application pursuant to 35 U.S.C. 111(a)** without a signed Declaration. Please accord a filing date and a serial number to such application and inform the undersigned thereof so that a signed Declaration and the surcharge required by 37 C.F.R. 1.16(e) may be duly filed.

Please address all correspondence to:

Thomas J. Kowalski, Esq.  
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Respectfully,



Thomas J. Kowalski  
Registration No. 32,147  
Attorney for Applicants  
Enclosure

JMM3533

**IN THE UNITED STATES PATENT AND TRADEMARK OFFICE**

Applicant(s) : Philip Michael SAVAGE  
U.S. Serial No. : Filed Concurrently Herewith  
Int'l Appln. No. : PCT/GB99/01764  
Int'l Filing Date : 04 June 1999  
Earliest Priority Date : 05 June 1998  
For : METHOD FOR PRODUCING OR ENHANCING A  
T-CELL RESPONSE AGAINST A TARGET CELL  
USING A COMPLEX COMPRISING AN HLA  
CLASS 1 MOLECULE AND AN ATTACHING  
MEANS

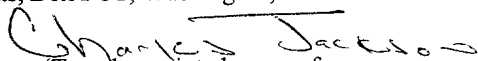
745 Fifth Avenue  
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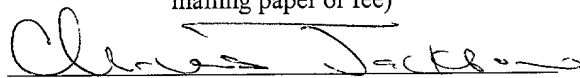
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**PRELIMINARY AMENDMENT**

Assistant Commissioner for Patents  
Box PCT  
Washington, D.C. 20231

Dear Sir:

Preliminary to the examination of this U.S. application, please enter the following  
amendments:

**IN THE SPECIFICATION:**

Page 1, below "METHOD FOR PRODUCING OR ENHANCING A T-CELL RESPONSE AGAINST A TARGET CELL USING A COMPLEX COMPRISING AN HLA CLASS 1 MOLECULE AND AN ATTACHING MEANS" please insert:

**--RELATED APPLICATIONS**

This application is the continuation-in-part of PCT/GB99/01764, filed June 4, 1999, designating the U.S. and published as WO 99/64464, with claims of priority from Great Britain application nos. 9812227.8, filed June 5, 1998 and 9908333.9 filed April 12, 1999. All of the foregoing applications, as well as all documents cited in the foregoing applications ("application documents") and all documents cited or referenced in application documents are hereby incorporated herein by reference. Also, all documents cited in this application ("herein cited documents") and all documents cited or referenced in herein cited documents are hereby incorporated herein by reference.

**IN THE CLAIMS:**

Claim 4, line 1, please delete "or claim 3";

Claim 5, line 1, please delete "or claim 3";

Claim 9, line 1, please delete "any preceding claim" and insert --claim 1--;

Claim 10, line 1, please delete "any preceding claim" and insert --claim 1--;

Claim 11, line 1, please delete "any preceding claim" and insert --claim 1--;

Claim 12, line 1, please delete "appended to claim 10";

Claim 12, line 1, please delete "said" and insert --there is a--;

Claim 12, line 2, after "peptide" please insert --that--;

Claim 13, lines 1-2, please delete "to claim 10, or claim 12 wherein said" and insert --there is a--;

Claim 13, line 2, after “peptide” insert --that--;

Claim 14, line 1, please delete “ay of claims 11-13” and insert --claim 11--;

Claim 15, line 1, please delete “any of claims 1-10” and insert --claim 1--;

Claim 16, line 1, please delete “appended to claim 10, wherein said” and insert --there is

a--

Claim 16, line 2, after “peptide” insert --that--;

Claim 17, line 1, please delete “any preceding claim” and insert --claim 1--;

Claim 18, line 1, please delete “any of claims 1-16” and insert --claim 1--;

Claim 19, line 1, please delete “any of claims 1-18” and insert --claim 1--;

Claim 20, line 1, please delete “or claim 16”;

Claim 21, line 2, please delete “any of claims 1-18” and insert --claim 1--;

Claim 21, line 3, please delete “in accordance with the method of claim 19” and insert  
--by introducing to said target cell a HLA class 1 molecule or fragment thereof and

attaching means--.

Claim 22, lines 7-8, please delete “or claim 16”;

Claim 24, lines 7-8, please delete “or claim 16”;

Claim 25, line 6, please delete “any of claims 11-14” and insert --claim 11--;

Claim 26, line 6, please delete “any of claims 11-14” and insert --claim 11--;

Claim 27, line 1, please delete “any of claims 11-14” and insert --claim 11--; and

Claim 28, line 2, please delete “or claim 26”.

**IN THE ABSTRACT:**

Please add the Abstract attached hereto as a separate sheet.

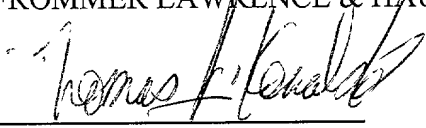
REMARKS

This application includes multiple claim dependencies. The amendment removes the multiple claim dependencies, and the filing fee for this application was computed on the basis that no dependent claim depends from more than one preceding claim.

Entry of this amendment and an early examination on the merits are respectfully solicited.

Respectfully submitted,  
FROMMER LAWRENCE & HAUG LLP

By:

  
Thomas J. Kowalski  
Reg. No. 32,147  
(212) 588-0800

008211 5864260

METHOD FOR PRODUCING OR ENHANCING A T-CELL RESPONSE AGAINST A TARGET CELL USING A COMPLEX COMPRISING AN HLA CLASS I MOLECULE AND AN ATTACHING MEANS

- 5 This application relates to means for producing or enhancing an immunological response against a target cell, through the attachment of an immunogenic HLA class I molecule thereto. The invention is of potential use in the prevention and treatment of malignant diseases including cancer and leukaemia, infectious diseases including viral infections such as HIV, bacterial infections including
- 10 tuberculosis, and parasitic infections including malaria.

- Cytotoxic T cells in the cellular immune system are responsible for recognising cells that display "foreign" markings, and triggering an immunological response against such cells. Each cytotoxic T cell expresses a number of cell surface
- 15 recognition receptors, which recognition receptors all possess precise specificity for a particular "foreign" peptide sequence, which recognition receptors are adapted to bind to HLA class I molecules expressed on the surface of cells scanned by the T cell. HLA class I molecules are cell surface molecules which possess a peptide binding groove exposed on the external surface of the cell,
- 20 which groove is arranged under normal circumstances to bind a peptide derived from the interior of the cell. When a recognition receptor on a cytotoxic T cell



binds to an HLA class I molecule on the surface of a scanned cell, the recognition receptor is enabled to contact the peptide binding groove of the HLA class I molecule and interact with any peptide contained therein. If this peptide matches the specificity of the recognition receptor, the T cell is said to

5 recognise the scanned cell, and may consequently trigger an immunological response against said scanned cell.

Cytotoxic T cells of various specificities within a host immune system are also able to recognise and trigger an immunological response against a cell

10 exhibiting an HLA class I molecule which is of a different allotype from the HLA class I molecules of the host cells. An immunological response of this kind is known as an "alloreactive" response.

An immunological response against a cell usually results in the lysis of the cell

15 and/or the local release of cytokines. It has however been observed that cytotoxic T cells do not trigger the lysis of so-called antigen presenting cells (APCs) in this way. Instead, the immunological response triggered by T cell recognition of an HLA class I molecule on the surface of an antigen presenting cell results in the direct selective proliferation of the cytotoxic T cell. The host

20 immune system consequently becomes immunised against any cells exhibiting the foreign peptide recognised by the surface recognition receptors on this T

cell.

It is recognised that the effector mechanisms of the cellular immune system could be a powerful tool in the prevention and treatment of many illnesses, including malignant processes and infectious and auto-immune diseases, including cancer. A small number of the HLA class I molecules on a tumour cell surface may be found to bind peptides which are selectively expressed or over-expressed in tumour cells and are capable of being recognised by cytotoxic T cells in the immune system. Such peptides may furthermore be tumour specific, being found only infrequently, or not at all, on the HLA class I molecules of non-tumour cells. An example of one such tumour specific peptide is the HMW-MAA antigen found on melanoma cells. However, the number of HLA molecules presenting such peptides is generally too small to stimulate an effective immunological response against the tumour cell. Moreover, such peptides are rarely, if ever, presented by HLA class I molecules on the surface of APCs.

Attempts to enhance the response of the cellular immune system to tumour cells have hitherto focused on increasing tumour cell immunogenicity. In particular, various efforts have been made to produce high-level expression of immunogenic HLA class I molecules on the surface of tumour cells, through

the techniques of gene therapy. The delivery of cDNA encoding an HLA class I gene containing an immunogenic peptide in the leader sequence of the HLA molecule has been described in Kang (Cancer Res. 57, 1997, 202-205).

Meanwhile, Stopeck (J Clinical Oncology 15, 1997, 341-349) describes the

5 transfection of allogeneic HLA class I in patients with melanoma. This work has demonstrated some response in clinical trials, but has also highlighted the difficulties involved in targeting tumour cells at multiple sites in vivo through the techniques of gene therapy.

10 The present application sets out to provide improved means for producing or enhancing an immunological response against a target cell, and to provide an improved method for treating or preventing cancer and other malignant, infectious or auto-immune diseases.

15 Accordingly, in one aspect of the present invention there is provided a complex comprising an HLA class I molecule or fragment thereof, which HLA class I molecule or fragment thereof comprises a T cell binding portion, and attaching means for selectively attaching said HLA class I molecule or fragment thereof

20 to a target cell, characterised in that said HLA class I molecule or fragment thereof binds or is attached to a peptide, which recognition peptide is arranged to be presented by said HLA class I molecule or fragment thereof.  
In another aspect of the present invention there is provided a method of

attaching an HLA class I molecule or fragment thereof to a target cell, which HLA class I molecule or fragment thereof comprises a T cell binding portion, comprising the step of introducing to said target cell said HLA class I molecule or fragment thereof and attaching means for selectively attaching said HLA

5 class I molecule or fragment thereof to the target cell.

In yet another aspect of the present invention, there is provided a pharmaceutical composition comprising an HLA class I molecule or fragment thereof, which HLA class I molecule or fragment thereof comprises a T cell

10 binding portion; attaching means for selectively attaching said HLA class I molecule or fragment thereof to a target cell; and an appropriate excipient or carrier.

The HLA class I molecule or fragment thereof may bind a peptide, which

15 peptide is arranged to be presented for T cell recognition by said HLA class I molecule or fragment thereof. Said peptide may be attached to the HLA class I molecule or fragment thereof in accordance with the method described in Garboczi (PNAS 89, 1992, 3429-3433).

20 The attaching means preferably comprises a linking polypeptide with high specific affinity for a target cell specific molecule on the surface of the target

cell. By "target cell specific molecule" herein is meant any molecule that is characteristically expressed or over-expressed on the surface of the target cell.

By way of example, in cancer cells said "target cell specific molecule" could include any of the following tumour associated antigens: carcinoembryonic

5 antigen, placental alkaline phosphatase, polymorphic epithelial mucin, human chorionic gonadotrophin, CD20, prostate specific antigen, ca-125, HMW-MAA and others.

Conveniently, the linking polypeptide will comprise an antibody, preferably a  
 10 monoclonal antibody, raised against said target cell specific molecule (Riethmuller and Johnson, Curr. Opin. Immunol. 4, 1992, 647-655). Suitable antibodies for this purpose include C46, 85A12, H17E2, HMFG1, W14, 1F5, 225.28s (Buraggi 1985 Cancer Res. 45, 3378-3387), and others. Deposits of the immortalised hybrids producing these antibodies have been made at the  
 15 American Type Culture Collection, Rockville MD, USA. Further examples of antibodies are described in Maloney et al (Blood 84, 1994, 2457-2466), Riethmuller et al (Lancet 343, 1994, 1177-1183) and Hird et al (Br. J. Cancer 68, 1993, 403-406).

20 Said linking polypeptide may comprise an antibody raised against a target cell specific molecule and a coupling system for coupling said antibody to said

HLA class I molecule or fragment thereof. The coupling system may comprise a two- or three-step chain of well-characterised paired small molecules, joined to the antibody and the HLA class I molecule so as to form a stable bridge between the two. Examples of paired small molecules which might be used in this connection include (but are not limited to) biotin and avidin/streptavidin (Moro, 1997 Cancer Res. 57, 1922-1928; Altman et al, Science 274, 1996, 94-96), and calmodulin and calmodulin binding peptides (Neri, 1996, J. Invest. Dermatol. 107, 164-170). Alternatively, said linking polypeptide may comprise an antibody-raised against a target cell specific molecule, which antibody is adapted to be attached directly to said HLA class I molecule or fragment thereof.

In a further possible embodiment of the invention, said complex may comprise a recombinant protein, which recombinant protein includes a moiety comprising said HLA class I molecule or fragment thereof, and a moiety comprising said attaching means.

The HLA class I molecule or fragment thereof may be purified from plasma or platelets or made recombinantly. The HLA class I molecule or fragment thereof may further be arranged to bind and present for T cell recognition a defined peptide of choice, such as a viral, bacterial, parasitic, or tumour-specific

10

Advantageously, the target cell is a tumour cell or any diseased or foreign cell the presence of which is undesired in a patient, such as a cancer cell,

leukaemia cell, a cell infected with the HIV virus or with any other microbe or virus, a cell responsible for detrimental activity in auto-immune disease, and so on. In order to accelerate the triggering of an immunological response against said target cell in a patient, said HLA class I molecule or fragment thereof will preferably be capable of producing a powerful immune response from the cellular immune system of the patient. Accordingly, said HLA class I molecule or fragment thereof may bind a viral or microbial peptide, preferably a viral or

microbial peptide to which the patient is likely to have had previous exposure. In particular, said HLA class I molecule or fragment thereof may bind an influenza virus peptide, a measles virus peptide, an Epstein-Barr virus peptide, in particular an Epstein-Barr virus peptide comprising the RAKFFQLL epitope of the lytic protein BZLF1, a Cytomegalovirus peptide, or a tetanus toxoid peptide. Alternatively, said HLA class I molecule or fragment thereof may bind any peptide which already has a strong cytotoxic T cell response or which is capable of inducing a powerful immune response. The allotype of said HLA class I molecule or fragment thereof may additionally ~~or alternatively~~ be different from the allotype of the HLA class I molecules of the patient, so that an alloreactive response may additionally ~~or alternatively~~ be triggered against said target cell.

In another embodiment of the invention the target cell is an antigen presenting cell (APC). Recognition by a cytotoxic T cell of an HLA class I molecule or fragment thereof attached to said APC may result in direct and selective proliferation of the cytotoxic T cell. Accordingly, said HLA class I molecule or fragment thereof will advantageously be adapted to present for T cell recognition a tumour specific peptide as defined above, or a viral peptide, or a bacterial peptide, or a parasitic peptide, or any peptide which is exclusively or characteristically presented by HLA class I molecules on the surface of



- diseased, malignant or foreign cells the presence of which is undesirable in a patient. Peptides linked to malignant conditions have been characterised (Brossart, 1998 Cancer Res. 58, 732-736 and Lucas, 1998 Cancer Res. 58, 743-752), as have peptides of parasitic origin (Khusmith, 1991 Science 252, 715-718)
- 5 718) The attachment of an HLA class I molecule or fragment thereof to an APC, in accordance with the present invention, may be used for in vivo immunisation against cells presenting a given peptide, or ex vivo production of cytotoxic T cells of a particular specificity.
- 10 Where the target cell is a tumour cell or microbially infected cell, the pharmaceutical composition of the present invention may be used for the treatment of a tumour or microbial disease respectively, and there is provided a method of treating a tumour or microbial disease in a patient, comprising the step of administering to a patient in need thereof an effective amount of said
- 15 pharmaceutical composition.

It must be noted that whilst many tumour types express tumour associated antigens, heterogeneity in the level of expression does occur, so some tumour cells may not be targeted by antibody and lysed directly. However, in vitro data

20 from the analogous antibody-superantigen system shows that the high local levels of cytokines released by activated T cells can lead to the death of

untargeted bystander tumour cells (Dohlsten et al, Int. J. Cancer 54, 1993, 482-488) It is likely that similar effects will occur in a targeting system using MHC class I/peptide complexes. Similarly, it is possible that the presence of activated cytotoxic T cells releasing cytokines in the tumour may lead to

5 enhancement of a specific anti-tumour immune response.

Where the target cell is an APC and the HLA class I molecule or fragment thereof binds a tumour-specific peptide or any peptide which is exclusively or characteristically presented by HLA class I molecules on the surface of a virally, bacterially, parasitically or microbially infected cell, the pharmaceutical

10 composition of the present invention may be used for immunising against the tumour or viral, bacterial, parasitic or microbial infection respectively, and there is provided a method of immunising against a tumour or viral, bacterial, parasitic or microbial infection in a patient, comprising the step of administering to a patient in need thereof an effective amount of said

15 pharmaceutical composition.

The response of said patient may be improved by in vivo cytokine support, or by the infusion of antigen-specific cytotoxic T cells expanded ex vivo.

Transient immunosuppression (Ledermann et al, Int. J. Cancer 47, 1991, 659-664) may be used to minimise the immunogenic response of a patient to components of the targeting system such as the avidin bridge.

20

The administration of said pharmaceutical composition may be by way of oral, sublingual, transdermal or parenteral administration.

Said effective amount of the pharmaceutical composition will depend on factors  
5 such as the nature and severity of the disorder being treated and on the weight, age and condition of the patient.

For oral or parenteral administration, it is greatly preferred that the pharmaceutical composition is administered in the form of a unit-dose  
10 composition, such as a unit dose oral or parenteral composition.

Such compositions are prepared by admixture and are suitably adapted for oral or parenteral administration, and as such may be in the form of tablets, capsules, oral preparations, powders, granules, lozenges, reconstitutable  
15 powders, injectable and liquid infusible solutions or suspensions or suppositories.

Tablets and capsules for oral administration are usually presented in a unit dose, and contain conventional excipients such as binding agents, fillers,  
20 diluents, tableting agents, lubricants, disintegrants, colourants, flavourings, and wetting agents. The tablets may be coated according to well known methods in

the art.

Suitable fillers for use include cellulose, mannitol, lactose and other similar agents. Suitable disintegrants include starch, polyvinylpyrrolidone and starch derivatives such as sodium starch glycolate. Suitable lubricants include, for example, magnesium stearate. Suitable pharmaceutically acceptable wetting agents include sodium lauryl sulphate

These solid oral compositions may be prepared by conventional methods of blending, filling or tableting. Repeated blending operations may be used to distribute the active agent throughout those compositions employing large quantities of fillers. Such operations are, of course, conventional in the art.

Oral liquid preparations may be in the form of, for example, aqueous or oily suspensions, solutions, emulsions, syrups, or elixirs, or may be presented as a dry product for reconstitution with water or other suitable vehicle before use. Such liquid preparations may contain conventional additives such as suspending agents, for example sorbitol, syrup, methyl cellulose, gelatin, hydroxyethylcellulose, carboxymethyl cellulose, aluminium stearate gel or hydrogenated edible fats, emulsifying agents, for example lecithin, sorbitan monooleate, or acacia; non-aqueous vehicles (which may include edible oils),

for example, almond oil, fractionated coconut oil, oily esters such as esters of glycerine, propylene glycol, or ethyl alcohol; preservatives, for example methyl or propyl p-hydroxybenzoate or sorbic acid, and if desired conventional flavouring or colouring agents.

5

Oral formulations also include conventional sustained release formulations, such as tablets or granules having an enteric coating.

For parenteral administration, fluid unit dose forms may be prepared  
10 comprising a sterile vehicle. The components of the composition, depending on the vehicle and the concentration, can be either suspended or dissolved. Parenteral solutions are normally prepared by dissolving the components of the composition in a vehicle and filter sterilising before filling into a suitable vial or ampoule and sealing. Advantageously, adjuvants such as a local anaesthetic,  
15 preservatives and buffering agents are also dissolved in the vehicle. To enhance the stability, the composition may be frozen after filling into the vial and the water removed under vacuum.

Parenteral suspensions are prepared in substantially the same manner except  
20 that the compound may be suspended in the vehicle instead of being dissolved and sterilised by exposure to ethylene oxide before suspending in the sterile

vehicle. Advantageously, a surfactant or wetting agent may be included in the composition to facilitate uniform distribution of the compound of the invention.

As is common practice, the compositions will usually be accompanied by

5 written or printed directions for use in the treatment concerned

Following is a description, by way of example only, and with reference to the accompanying drawings, of methods of putting the present invention into effect.

10 In the drawings :-

Figure 1 shows a diagram showing the method/idea for delivering HLA molecules to the surface of tumour cells.

15 Figure 2 shows a FACs analysis of HLA-A2-ve Mel 2 melanoma cells treated with biotin-conjugated monoclonal antibody 225.28s, avidin, biotin-conjugated HLA-A2/gag complexes, anti-HLA-A2 monoclonal antibody BB7.2 and phycoerythrin-conjugated rabbit anti-mouse antibody.

20 Figure 3 shows the results of a T cell cytotoxicity chromium release assay with Mel 1 cells treated with the delivery system of biotin-conjugated monoclonal

antibody 225.28s, avidin, and biotin-conjugated HLA-A2/gag complexes. These cells were incubated with HLA-A2/gag specific cytotoxic T cells with effector/target ratios of 0:1 - 20:1 for 20 hours.

- 5 Figure 4 shows a diagram showing the method/idea for delivering HLA class I/peptide complexes to antigen presenting cells.

- Figure 5 shows the results of an ELISA assay, described in Example 2 below, for demonstrating the stability of various MHC class I/peptide complexes at  
10 37°C. The results shown are the mean of assays performed on each sample in triplicate.

- Figure 6 shows a FACS analysis of HLA-class I deficient Daudi cells targeted with HLA-A2 via biotinylated anti-CD20 mAb. Trace 1 (lefthand trace)  
15 corresponds to native untargeted Daudi cells. Trace 2 (righthand trace) corresponds to Daudi cells targeted with mAb/avidin/HLA-A2/gag/FITC anti-MHC class I. Mean fluorescence trace 1 = 0.31, mean fluorescence trace 2 = 24.3 (arbitrary fluorescence units).

- 20 Figure 7 shows the results of a four hour chromium release assay, described in Example 2 below, in which HLA class I-deficient Daudi cells targeted with

various components of the HLA-A2/gag delivery system were incubated with HLA-A2/gag specific cytotoxic T cell clones. A comparison was made with native and peptide-pulsed 221.A2 cells (HLA-A2 +ve).

- 5 Figure 8 shows the results of a four hour chromium release assay, described in Example 2 below, in which HLA-A2/gag targeted Daudi cells were incubated with HLA-A2/gag-specific and HLA-A2/Melan A-specific cytotoxic T cell clones.
- 10 Figure 9 shows the results of a twenty hour chromium release assay, described in Example 2 below, in which HLA-A2 +ve SK29.Mel cells were incubated with HLA-A2/gag specific cytotoxic T cell clones.

#### Example 1

15

The following components were used:

- 20 Target cells : A human melanoma cell line Mel 1, deposited at the Department of Immunology, Institute of Molecular Medicine, Oxford, that carries the HLA class I allotype HLA-A2. The cell line was grown in



A human melanoma cell line Mel 2, deposited at the Department of Immunology, Institute of Molecular Medicine, Oxford, that does not carry the HLA class

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molecules, as described in Altman 1996, Science 274, 94-96, further containing the "gag" peptide that is part of the HIV virus. This peptide comprises the amino acid sequence -SLYNTVATL-. Methods for the preparation/isolation thereof are described in Johnson 1991, J Immunol

20

T cells : HLA-A2/gag specific cytotoxic T cells obtained from an A2+ve HIV patient as described in Altman 1994, Science 274, 94-96.

The binding of recombinant HLA-A2 to the treated Mel 2 cells was shown by the attachment of anti-HLA-A2 monoclonal antibody BB7.2 (Santos-Aguado 1988, J. Immunol 141, 2811-2818) following incubation with BB7.2 antibody at a final concentration of 10µg/ml at 37°C for 30 minutes. After washing in

tissue culture media the cells were incubated with phycoerythrin conjugated rabbit anti-mouse antibody (Sigma, Poole, UK) at a final concentration of 10µg/ml for 30 minutes at 37°C and analysed in a Becton Dickson Facscan machine. The result of this analysis is shown in Fig 2 which demonstrates a  
5 positive signal indicating the presence of HLA-A2 molecules attached to the surface of the Mel 2 cells.

A chromium release T cell cytotoxicity assay was then performed in order to establish the ability of HLA-A2/gag specific T cell clones to lyse Mel 1 cells  
10 coated with HLA-A2/gag in accordance with the present method.

Approximately  $10^6$  Mel 1 cells were first pre-incubated with 1.85µBq  $\text{Na}_2^{51}\text{CrO}_4$  (obtained from Amersham International, Amersham, UK) for 1 hour at 37°C. The pre-incubated Mel 1 cells were then incubated with biotin conjugated monoclonal antibody 225.28s at a final concentration of 20µg/ml at  
15 37°C for 30 minutes, and washed in tissue culture media. Following this, the Mel 1 cells were incubated with avidin at a final concentration of 10µg/ml for 10 minutes at 37°C and then washed again in tissue culture media. The Mel 1 cells were then incubated with biotin conjugated HLA class I HLA-A2/gag molecules at a final concentration of 20µg/ml at 37°C for 20 minutes and  
20 washed with tissue culture media.

Having been coated with HLA class I HLA-A2/gag, the chromium-treated Mel 1 cells were then incubated with HLA-A2/gag specific cytotoxic T cells in ratios of 0:1 to 20:1 of effector to target cells at 37°C for 20 hours. Lysis of Mel 1 cells treated with  $\text{Na}_2^{51}\text{CrO}_4$  results in the release of radioactive chromium, which may be detected by analysis in a scintillation counter.

In order to establish the percentage of Mel 1 cells lysed following incubation with HLA-A2/gag specific cytotoxic T cells, the following measurements were taken: background release of chromium from the Mel 1 cells in media alone ("M"); release of chromium from the Mel 1 cells following incubation with the T cells ("E"); release of chromium from the Mel 1 cells following final treatment with 5% Triton X-100 detergent ("T"). Treatment with detergent will cause the lysis of all the remaining intact Mel 1 cells.

% Mel 1 lysis by cytotoxic T cells was calculated as follows.

$$\% \text{ lysis} = 100 \times \frac{(E - M)}{(T - M)}$$

This analysis was carried out on Mel 1 cells treated with biotin-conjugated 225.28s, avidin, and biotin-conjugated HLA-A2/gag. As a control, the analysis was also carried out on Mel 1 cells treated with biotin-conjugated 225.28s and

The key results of this analysis are illustrated in Figure 3, which indicates that significant lysis (20%) of Mel 1 cells by HLA-A2/gag specific cytotoxic T cells occurs only when the Mel 1 cells have been treated with all the components of the attaching and delivery means of the present invention (ie biotin-conjugated 225.28s monoclonal antibodies, avidin, and biotin-conjugated HLA-A2/gag) No significant increase in cell lysis over background levels was observed in either of the control runs.

The following components were used:

Target cells : The Daudi B cell line (MHC class I-negative) melanoma line SK-mel-29 (HLA-A2.1-positive), .221/A2 (HLA-A2.1-positive), were maintained in RPMI media with 10% fetal calf serum and antibiotics in a 37°C incubator with 5% CO<sub>2</sub>.

Attaching means : Monoclonal antibodies 225.28s (Buraggi 1985  
Cancer Res. 45, 3378-3387) and 2H7 that bind to the  
HMW-MAA antigen. Biotin is chemically conjugated  
onto these antibodies as described in Bayer 1990,  
5 Methods Embryology 184, 138-160.

Pure hen egg avidin obtained commercially from  
Societa Prodotti Antibiotici, Milan, Italy.

10 HLA : Biotinylated complexes of recombinant MHC class I  
and peptide were produced as described previously  
(Altman et al, Science 274, 1996, 94-96; Ogg et al,  
Science 279, 1998, 2103) Prokaryotic expression of  
B2M and MHC class I heavy chain, modified by the  
15 C terminal addition of a target sequence for the  
biotin ligase enzyme BirA, was followed by  
inclusion body purification. Following refolding of  
heavy chain and B2M around specific peptide,  
complexes of 45kD were isolated by gel filtration,  
20 biotinylated overnight using BirA in the presence of  
ATP, Mg<sup>2+</sup> and biotin, and then purified by gel

filtration and anion exchange.

T cells : Human cytotoxic T cell clones 010 (specific for  
HLA-A2/gag 77-85 = SLYNTVATL (Parker et al, J  
5 Immunol. 149, 1992, 3580-3587)) and IF9 (specific  
for HLA-A2/melan-A 26-35 = EAAGIGILTV  
(Romero et al, J Immunol 159, 1997, 2366) were  
maintained in media supplemented with 5% human  
serum and IL-2 100 IU/ml.

10

The stability of the MHC class I/peptide complexes was first established by an  
ELISA assay. Various MHC class I/peptide complexes, including HLA-  
A2/Gag3Y, HLA-A2/Gag3F, HLA-A2/Lmp2, HLA-B35/Env and HLA-B35/nef,  
were prepared as outlined above, and were pre-incubated at 10ug/ml in tissue  
15 culture media for 0-20 hours at 37°C. ELISA plates were coated with the mAb  
W6/32 (5ug/ml in carbonate buffer pH 9.6 overnight at 4°C) which recognises  
conformationally correct MHC class I molecules (Parham, 1979), and then  
blocked by incubation in 1% bovine serum albumin for 2 hours at 37°C. The  
MHC class I/peptide complexes were incubated for 30 minutes with the ELISA  
20 plates at room temperature, and binding was detected with rabbit anti-human  
B2 microglobulin followed by alkaline phosphatase conjugated goat anti-rabbit

immunoglobulin, and substrate. All incubations were separated by extensive washes in PBS.

Absorbances at 600nm were measured in a Titertek Multiscan ELISA reader.

5 Three assays were performed for each sample, and the mean reading was calculated.

The results obtained with samples preincubated for 0, 1, 4, 16 and 20 hours are shown in Figure 5. The results demonstrate that the HLA-A2/gag complexes  
10 have appreciable stability in culture media at 37°C, with an estimated half-life in excess of 24 hours.

In storage at 0.5-lmg/ml at 4°C HLA-A2/gag complexes appear to be stable for at least 12 months (data not shown).

15

To demonstrate the ability of the attaching means to cause display of MHC class I on the surface of Daudi cells, Daudi cells deficient in MHC class I were sequentially incubated at 4°C with biotinylated anti-CD20 (Ancell, Nottingham, UK; mAb 2H7 (Berenson et al, Blood 67, 1986, 509-515) at 1ug/ml for 30  
20 minutes); hen egg avidin (S.P.A., Milan, Italy, at 10ug/ml for 10 minutes); biotinylated HLA-A2/gag (at 10ug/ml for 10 minutes); and FITC labelled



anti-MHC class I (Ancell, Nottingham, UK; mAB 3F10 (Eisenbarth et al, J. Immunol. 124, 1980,1237-1244) at 10 ug/ml). Parallel controls omitted one or other incubation. Cells were washed 3 times in PBS between stages and then fixed in PBS plus 2% formaldehyde and analysed by flow cytometry.

5

Cells incubated with all three layers of the labelling system had high levels of detectable MHC class I/peptide on their surface compared to untreated Daudi cells (Figure 6). Cells treated with only any 2 components of the 3-step system gave fluorescence levels comparable to untreated cells (data not shown).

10

A chromium release cytotoxicity assay was carried out to establish the ability of specific T cell clones to lyse Daudi or SK-mel-29 cells in accordance with the present invention. Daudi or SK-mel-29 cells were incubated with  $^{51}\text{CrO}_4$  at 2uCi/uL for 1 hour at 37°C and then sequentially incubated with: the

15 biotinylated mAbs 2H7 or 225.28s (anti-HMW-MAA) respectively; avidin; and biotinylated HLA-A2/gag complexes as detailed above. Peptide pulsed target cells were incubated with gag 77-85 or melan-A 26-35 peptides at 0.1uM for 1 hour at 37°C. After washing, labelled target cells were plated into 96-well round bottom plates at 2,500 cells per well, followed by human CTLs at  
20 various effector:target ratios. Following incubation at 37°C, 20ul of supernatant was collected and the amount of  $^{51}\text{Cr}$  released was determined. The percentage

of cytotoxicity (lysis) obtained at each effector:target ratio was calculated as:  
 $100 \times (E-M)/(T-M)$ , where E = Experimental release, M = Release in media  
and T = Release in 5% Triton X-100 detergent.

- 5 The results shown in Figure 7 are the mean of experiments performed in  
duplicate. As shown by these results, the CTL clone (010) efficiently lysed  
HLA-A2-positive targets (.221/A2) only when these were pre-incubated with  
the HLA-A2/gag peptide. MHC class I-negative Daudi cells, when targeted  
with the HLA-A2/gag complexes of the present invention, were recognised and  
10 lysed by this CTL clone to an equivalent degree. Untargeted Daudi cells and  
cells targeted with only 2 of the 3 components of the targeting system were not  
recognised (maximal lysis <4% at E:T ratios of up to 80:1).

- Control CTL, showing a different HLA-A2-restricted specificity (HLA--  
15 A2/melan-A), did not lyse Daudi cells targeted with the HLA-A2/gag  
complexes (Fig. 8), demonstrating the fine specificity of the targeting approach.

- Untreated Daudi cells pulsed with gag peptide alone were not lysed by clone  
010 (data not shown), in keeping with their lack of endogenous MHC class I.

20

The ability of antibody-directed HLA-A2/gag complexes to sensitise the

melanoma cell line SK-mel-29 to lysis by HLA-A2/gag-specific CTL line is shown in Figure 8. At all E:T ratios, melanoma cells targeted by complexes linked to surface proteins were lysed substantially more than controls exposed to only two components of the 3-step targeting system. Additionally, MM9 melanoma cells that do not express HLA-A2 were also lysed in a similar manner (data not shown).

$$\begin{array}{c} \left\{ \begin{array}{c} 1 \\ 2 \\ 3 \\ 4 \\ 5 \\ 6 \\ 7 \\ 8 \\ 9 \\ 10 \\ 11 \\ 12 \\ 13 \\ 14 \\ 15 \\ 16 \\ 17 \\ 18 \\ 19 \\ 20 \\ 21 \\ 22 \\ 23 \\ 24 \\ 25 \\ 26 \\ 27 \\ 28 \\ 29 \\ 30 \\ 31 \\ 32 \\ 33 \\ 34 \\ 35 \\ 36 \\ 37 \\ 38 \\ 39 \\ 40 \\ 41 \\ 42 \\ 43 \\ 44 \\ 45 \\ 46 \\ 47 \\ 48 \\ 49 \\ 50 \\ 51 \\ 52 \\ 53 \\ 54 \\ 55 \\ 56 \\ 57 \\ 58 \\ 59 \\ 60 \\ 61 \\ 62 \\ 63 \\ 64 \\ 65 \\ 66 \\ 67 \\ 68 \\ 69 \\ 70 \\ 71 \\ 72 \\ 73 \\ 74 \\ 75 \\ 76 \\ 77 \\ 78 \\ 79 \\ 80 \\ 81 \\ 82 \\ 83 \\ 84 \\ 85 \\ 86 \\ 87 \\ 88 \\ 89 \\ 90 \\ 91 \\ 92 \\ 93 \\ 94 \\ 95 \\ 96 \\ 97 \\ 98 \\ 99 \\ 100 \end{array} \right\} \end{array}$$

## CLAIMS

- 1        A complex comprising an HLA class I molecule or fragment thereof,  
which HLA class I molecule or fragment thereof comprises a T cell binding  
5        portion, and attaching means for selectively attaching said HLA class I molecule  
or fragment thereof to a target cell.
- 2        A complex as claimed in claim 1, wherein said attaching means comprises  
a linking polypeptide with high specific affinity for a target cell specific molecule  
10        on the surface of the target cell.
- 3        A complex as claimed in claim 2, wherein said linking polypeptide  
comprises an antibody, preferably a monoclonal antibody, raised against said  
target cell specific molecule.
- 15        4        A complex as claimed in claim 2 or claim 3, wherein said linking  
polypeptide is adapted to be attached directly to said HLA class I molecule or  
fragment thereof.
- 20        5        A complex as claimed in claim 2 or claim 3, wherein said attaching means  
further comprises a coupling system for coupling said linking polypeptide to said

HLA class I molecule or fragment thereof.

6 A complex as claimed in claim 5, wherein said coupling system comprises  
a two- or three-step chain of well-characterised paired small molecules, which  
5 chain is joined to the linking polypeptide and the HLA class I molecule so as to  
form a stable bridge between the two

7 A complex as claimed in claim 6, characterised in that said chain  
comprises biotin and avidin/streptavidin

10

8 A complex as claimed in claim 6, characterised in that said chain  
comprises calmodulin and calmodulin binding peptide.

9 A complex as claimed in any preceding claim, which complex comprises  
15 a recombinant protein, which recombinant protein includes a moiety comprising  
said HLA class I molecule or fragment thereof, and a moiety comprising said  
attaching means.

10 A complex as claimed in any preceding claim, characterised in that said  
20 HLA class I molecule or fragment thereof binds or is attached to a recognition  
peptide, which recognition peptide is arranged to be presented by said HLA class

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I molecule or fragment thereof for T cell recognition.

11 A complex as claimed in any preceding claim, characterised in that said  
target cell is a type of cell the presence of which is undesirable in a patient, such  
5 as a tumour cell or a diseased, foreign or malignant cell such as a cancer cell, a  
leukaemia cell, a cell infected with the HIV virus or with any other parasite,  
bacterium, microbe or virus, or a cell responsible for detrimental activity in auto-  
immune disease.

10 12 A complex as claimed in claim 11 appended to claim 10, wherein said  
recognition peptide comprises a peptide which has a strong cytotoxic T cell  
response or which is capable of inducing a powerful immune response.

13 A complex as claimed in claim 11 appended to claim 10, or claim 12,  
15 wherein said recognition peptide comprises a viral or microbial peptide, such as  
an influenza virus peptide, a measles virus peptide, an Epstein-Barr virus peptide,  
in particular an Epstein-Barr virus peptide comprising the RAKFFQLL epitope  
of the lytic protein BZLF1, a Cytomegalovirus peptide, or a tetanus toxoid  
peptide.

20

14 A complex as claimed in any of claims 11-13, wherein the allotype of said

5     15     A complex as claimed in any of claims 1-10, wherein said target cell is an antigen presenting cell.

17 A complex as claimed in any preceding claim, wherein said target cell is  
15 a culture cell.

19 A method for attaching the complex of any of claims 1-18 to said target  
20 cell, comprising the step of introducing to said target cell said HLA class I





or auto-immune disease such as cancer or leukaemia, or an infectious disease such as a viral infection such as HIV infection, or a bacterial or microbial infection such as tuberculosis, or a parasitic infection such as malaria; said pharmaceutical composition comprising a complex as claimed in claim 15 or claim 16 and an appropriate excipient or carrier.

24 Use of the complex of claim 15 or claim 16 in the preparation of a medicament for use in immunising a patient against a disease or condition which is characterised by the presence in the patient's body of cells displaying said recognition peptide on the surface thereof; such as a tumour, or a malignant or auto-immune disease such as cancer or leukaemia, an infectious disease such as a viral infection such as HIV infection, a bacterial or microbial infection such as tuberculosis, or a parasitic infection such as malaria.

25 A method for the treatment of a disease or condition such as a tumour, or a malignant or auto-immune disease such as cancer or leukaemia, an infectious disease such as a viral infection such as HIV infection, a bacterial or microbial infection such as tuberculosis, or a parasitic infection such as malaria, comprising the step of administering to a patient in need thereof an effective amount of the complex of any of claims 11-14.

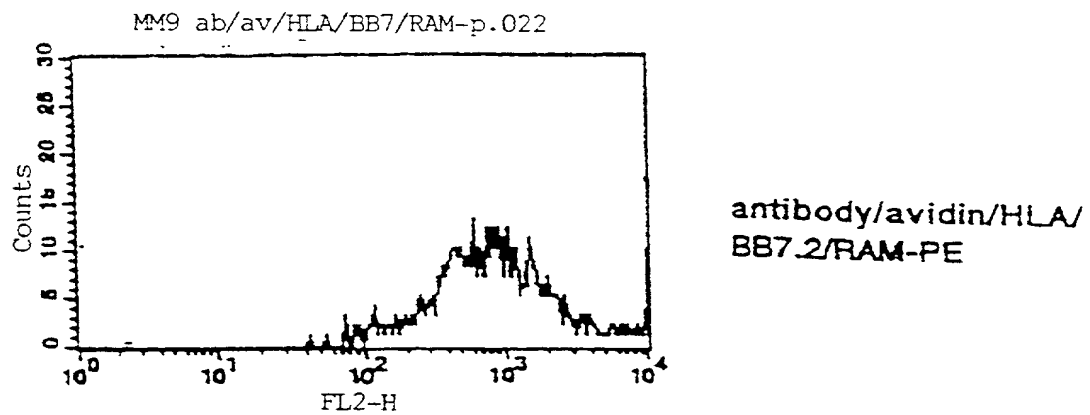
10

27 Use of the complex of any of claims 11-14 in the preparation of a medicament for the treatment of a tumour, or a malignant or auto-immune disease such as cancer or leukaemia, or an infectious disease such as a viral infection such as HIV infection, or a bacterial or microbial infection such as tuberculosis, or a parasitic infection such as malaria

20

A complex comprising an HLA Class 1 molecule and attaching means for selectively attaching the HLA class 1 molecule to a target cell is disclosed, and a method is provided for producing or enhancing an immunological response against a target cell, by attaching said complex to the target cell. Where the target is a diseased, foreign or malignant cell, this method may be used to promote the lysis of the targeted cell by T cells in the immune system. Where the target cell is an antigen presenting cell, this method may be used to promote the proliferation of specific T cell clones. The invention is of potential use in the prevention and treatment of malignant diseases including cancer and leukemia, infectious diseases including viral infections such as HIV, bacterial infections including tuberculosis, and parasitic infections including malaria.





MM9 melanoma 12.97

Figure 2

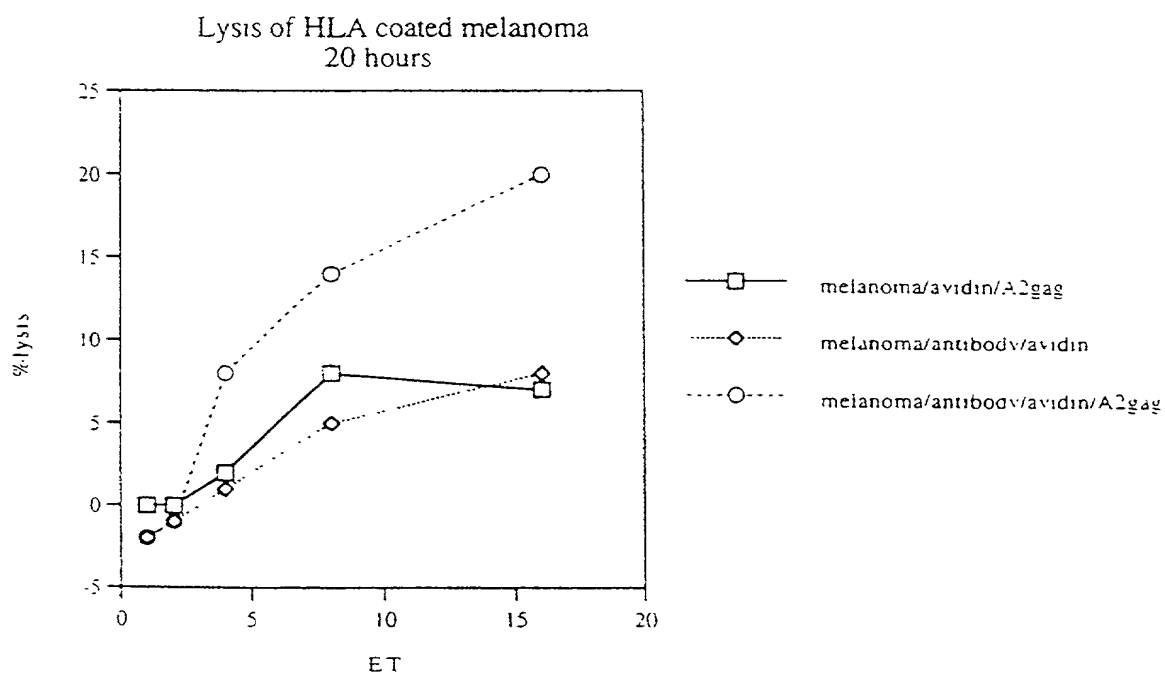
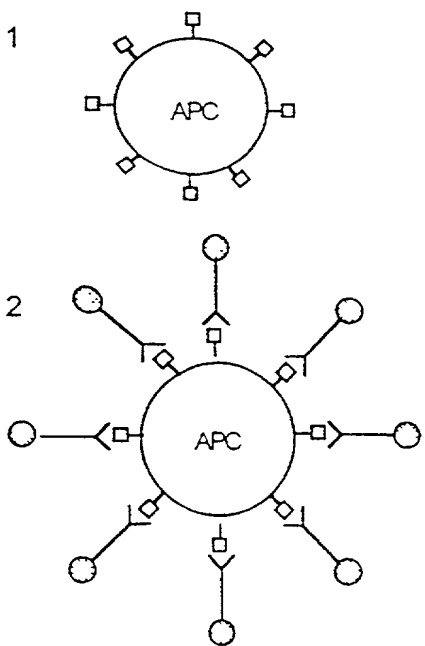




Figure 3




☐ APC Cell Surface Marker



Calmodulin linked  
monoclonal antibody



Calmodulin binding peptide linked HLA A2 with "tumour peptide"

 HLA A2 "tumour peptide" specific cytotoxic T cell

Naive T-cell

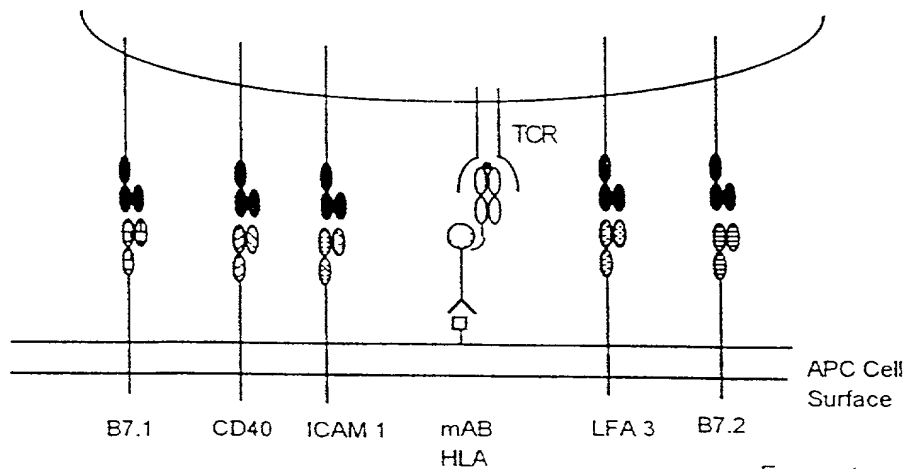


Figure 4

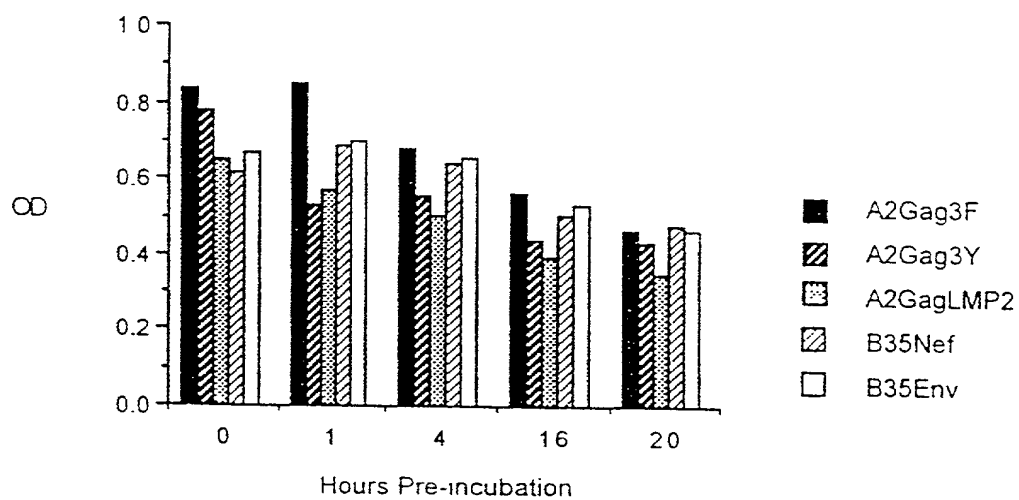


Figure 5



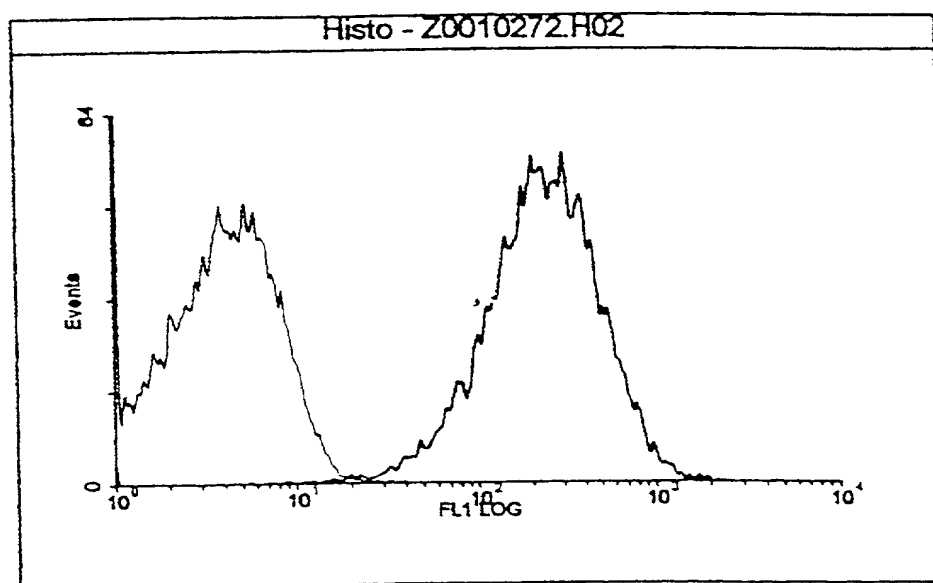


Figure 6

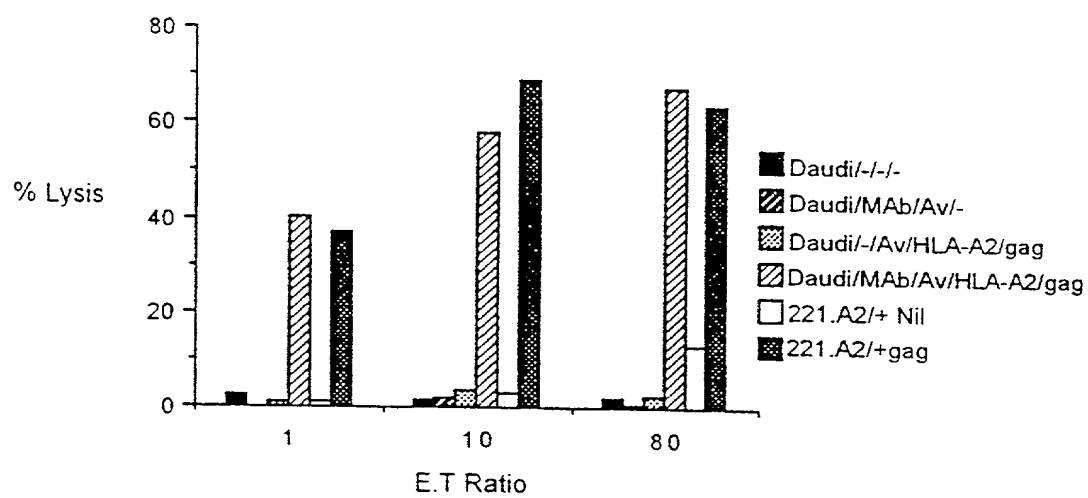


Figure 7

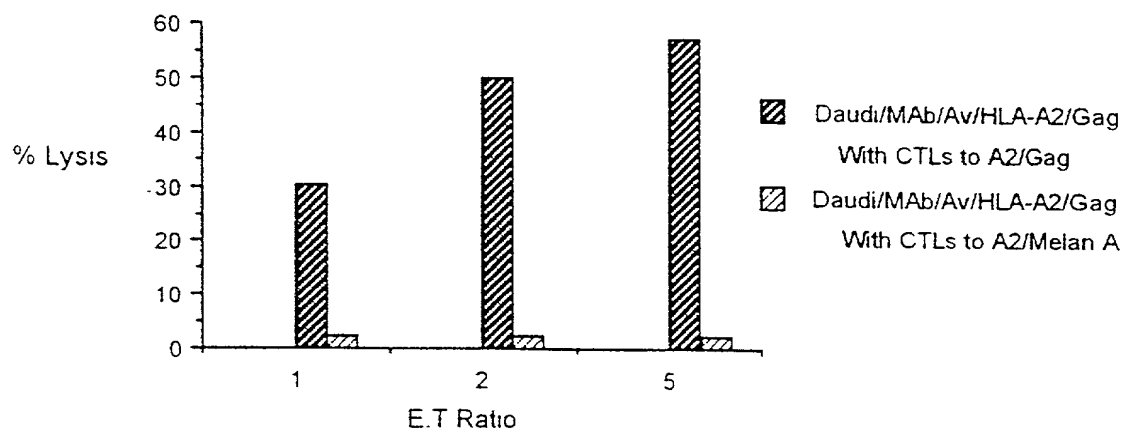


Figure 8

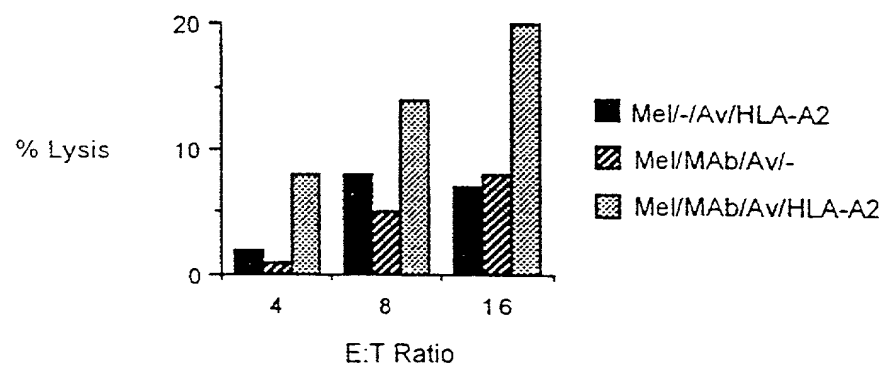


Figure 9

**DECLARATION FOR PATENT APPLICATION AND POWER OF ATTORNEY**  
(Includes reference to PCT International Applications)

FROMMER LAWRENCE & HAUG, LLP  
File No.: 674544-2001

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name.

I believe I am an original, first and joint inventor (if plural, names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention ENTITLED:

**METHOD FOR PRODUCING OR ENHANCING A T-CELL RESPONSE AGAINST A  
TARGET CELL USING A COMPLEX COMPRISING AN HLA CLASS I MOLECULE  
AND AN ATTACHING MEANS**

the specification of which:

is attached hereto

- X was filed on November 28, 2000 as:  
X United States Application Serial No. \_\_\_\_\_  
X as the National Phase or continuation or Divisional or Continuation-in-Part of  
PCT Application No. PCT/GB99/01764, filed JUNE 4, 1999 and designating the U.S., and  
published as WO 99/64464 on DECEMBER 16, 1999,  
X with amendments through DATE EVEN HEREWITH (if applicable, give details).

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose to the United States Patent and Trademark Office all information known to me to be material to patentability as defined in Title 37, Code of Federal Regulations, § 1.56.

I hereby claim foreign priority benefits under Title 35, United States Code § 119 (a) - (d) or § 365 (b) of any foreign application(s) for patent or inventor's certificate, or § 365 (a) of any PCT International application(s) designating at least one country other than the United State of America listed below and have also identified below any foreign application for patent or inventor's certificate or any PCT International applications designating at least one country other than the United States of America filed by me on the same subject matter having a filing date before that of the application(s) on which priority is claimed:

Prior Foreign/PCT Application(s) [list additional applications on separate page]:

Country (or PCT)

Application Number:

Filed (Day/Month/Year)

Priority Claimed:

Yes

No

Prior Foreign/PCT Application(s) [list additional applications on separate page]:

<u>Country (or PCT)</u>	<u>Application Number:</u>	<u>Filed (Day/Month/Year)</u>	<u>Priority Claimed:</u>	
			<u>Yes</u>	<u>No</u>
GREAT BRITAIN	9812227.8	05/JUNE/1998	X	
GREAT BRITAIN	9908333.9	12/APRIL/1999	X	

I hereby claim the benefit under 35 U.S.C. § 119(e) of any United States provisional application(s) listed below.

                     (Application Number)                           (Filing Date)

I hereby claim the benefit under Title 35, United States Code § 120 of any United States application(s) or § 365 (c) of any PCT international application(s) designating the United States of America that is/are listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in that/those prior United States or PCT International application(s) in the manner provided by the first paragraph of Title 35, United States Code § 112, I acknowledge the duty to disclose to the United States Patent and Trademark Office all information known to me to be material to patentability as defined in Title 37, Code of Federal Regulations, § 1.56 which became available between the filing date of the prior application and the national or PCT international filing date of this application:

Prior U.S. (or U.S.-designating PCT) Application(s) [list additional applications on separate page]:

U.S. Serial No.:    Filed (Day/Month/Year)    PCT Application No.    Status (patented, pending, abandoned)

04/JUNE/1999      PCT/GB99/01764      PUBLISHED AS WO 99/64464

I hereby appoint Thomas J. Kowalski, Registration No. 32,147, and FROMMER LAWRENCE & HAUG, LLP or their duly appointed associates, my attorneys or agents, with full power of substitution and revocation, to prosecute this application, to make alterations and amendments therein, to file continuation and divisional applications thereof, to receive the Patent, and to transact all business in the Patent and Trademark Office and in the Courts in connection therewith, and to insert the Serial Number of the application in the space provided above, and specify that all communications about the application are to be directed to the following correspondence address:

Direct all telephone calls to: (212) 588-0800  
to the attention of:  
Thomas J. Kowalski, Esq.

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Signature: \_\_\_\_\_ Date: \_\_\_\_\_

Post Office Address(es) of inventors [if different from residence]:

NOTE: In order to qualify for reduced fees available to Small Entities, each inventor and any other individual or entity having rights to the invention must also sign an appropriate separate "Verified Statement (Declaration) Claiming [or Supporting a Claim by Another for] Small Entity Status" form [e.g. for Independent Inventor, Small Business Concern, Nonprofit Organization, Individual Non-Inventor].